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2xYT Agar

Medium for fibrous bacteriophages

Practical information

Aplications

Preparation and recovery of competent cells

Categories Escherichia coli

Industry: Molecular biology / Microbiological Culture Media

Principles and uses

2xYT Agar is a nutritionally rich growth medium recommended for the optimal growth of recombinant strains of E. coli on agar plates. This medium is also used for propagation of M13 bacteriophage or other fibrous bacteriophages for sequencing and phage display.

Bacteriophages are viruses that can only infect and replicate within bacteria. In many cases, these are very specific relationships. Somatic coliphages specifically infect Escherichia coli and in water indicate contamination by human or animal faeces or by wastewater containing such material.

Tryptone and yeast extract are the sources for carbon, nitrogen, vitamins, minerals, and amino acids essential for growth, as well as growth factors that allow phages to reproduce without weakening the host cells. Sodium chloride supplies essential electrolytes for transport and osmotic balance. Many supplements, including antibiotics, are heat-sensitive and cannot be autoclaved. These should be filter-sterilized and added to the medium after it has cooled down and prior to solidification. Escherichia coli growth faster on the enrichment medium as provide aminoacid, nucleotide precursors, vitamins and other metabolite which in another way it should be synthesized by the cell.

Formula in g/L

Bacteriological agar	15	Sodium chloride	5
Tryptone	16	Yeast extract	10

Preparation

Suspend 46 grams of medium in one liter of distilled water. Mix well and dissolve by heating with frequent agitation. Boil for one minute until complete dissolution. Sterilize in autoclave at 121 °C for 15 minutes. Cool to 45-50 °C, mix well and dispense into plates.

Instructions for use

Inoculate and incubate at a temperature of 35±2 °C for 18-24 hours.

Quality control

Solubility	Appareance	Color of the dehydrated medium	Color of the prepared medium	Final pH (25⁰C)
w/o rests	Fine powder	Beige	Amber	7,0±0,2

Microbiological test

Incubation conditions: (35±2 °C / 18-24 h).

Microorganisms	Specification
Escherichia coli ATCC 23724	Good growth
Escherichia coli ATCC 33694	Good growth
Escherichia coli ATCC 33849	Good growth
Escherichia coli ATCC 39403	Good growth

Storage

2xYT Medium

For the cultivation of recombinant strains of E.coli and for growth of filamentous bacteriophages.

Practical information

Aplications Selective enrichment Preparation and recovery of competent cells

Industry: Culture media for Molecular biology

Principles and uses

2xYT Medium is a nutritive medium optimized for the growth and maintenance of M13 phages and other filamentous bacteriophages. It is also suitable for growth of recombinant strain of E.coli.

Categories

Escherichia coli

Escherichia coli

Tryptone provide nitrogen, vitamins, minerals and amino acids essential for growth. Yeast extract is source of vitamins, particularly the B-group. Sodium chloride supplies essential electrolytes for transport and osmotic balance. The components of the 2xYT Medium include nitrogen and other growth factors that allow bacteriophages to reproduce in large quantities without weakening the host. E. coli grows faster in this enriched medium, as it contains amino acids, precursors of nucleotides, vitamins and other metabolisms which otherwise the cell itself would have to synthesize.

Formula in g/L

Sodium chloride	5	Tryptone	16
Yeast extract	10		

Preparation

Suspend 31 grams of the medium in one liter of distilled water .Mix well and dissolve by heating with frequent agitation. Boil for one minute until complete dissolution. Dispense into appropriate containers and sterilize in autoclave at 121°C for 15 minutes.

Instructions for use

- Carry out the experimental procedure according to appropriate use or purpose.

- Inoculate and incubate at a temperature of 35±2 °C for 18-24 hours.

Quality control

Solubility	Appareance	Color of the dehydrated medium	Color of the prepared medium	Final pH (25°C)
w/o rests	Fine powder	Beige	Amber, slightly opalescent	7,0 ± 0,2

Microbiological test

Incubation conditions: (35±2 °C / 18-24 h)

Microorganisms

Escherichia coli ATCC 23724 Escherichia coli ATCC 33694 Escherichia coli ATCC 33849 Escherichia coli ATCC 39403 Escherichia coli ATCC 47014

Storage

Temp. Min.:2 °C Temp. Max.:25 °C Specification Good growth Good growth Good growth Good growth

2YT Autoinducible Growth Medium w/o Trace Elements

To grow IPTG inducible expression in bacterial strains.

Practical information

Aplications Protein expression Categories Escherichia coli

Industry: Culture media for Molecular biology

Principles and uses

Auto induction media was first formulated and developed by W. Studier to grow IPTG-inducible expression strains. The principle of auto induction media is based on carbon sources in the medium that are metabolized differentially to promote high density cell growth and automatically induce protein expression from lac promoters.

Auto induction media contains glucose as well as lactose as the carbon source. A limited concentration of glucose is metabolized preferentially during growth, which prevents uptake of lactose until the glucose is depleted, usually in mid to late log phase. As the glucose is depleted, lactose can be taken up and converted by the enzyme ß-galactosidase to the inducer allolactose. Allolactose causes the release of lac repressor from its specific binding sites in the DNA and thereby induces expression of T7 RNA polymerase from the lacUV5 promoter and unblocks T7lac promoters, allowing expression of target proteins by T7 RNA polymerase.

With Auto induction media, a high density cell growth is followed by a spontaneous induction of protein expression. There is no need to monitor the cell density and there is no conventional induction with IPTG. Parallel growth of many non-induced or auto-induced cultures is feasible because cultures are simply inoculated and grown to saturation. This is a great convenience and simplifies manual or automated induction and analysis of multiple clones compared to conventional IPTG induction, which requires monitoring growth of each culture and adding inducer at the proper stage of growth.

Formula in g/L

Glucose	0,5 A	Ammonium sulfate	3,3
Disodium phosphate	7,1 N	Aagnesium sulfate	0,15
Monopotassium phosphate	6,8 T	ryptone	16
Yeast extract	10 A	Alpha lactose	2

Preparation

Suspend 45,85 grams of the medium in one liter of distilled water. Mix well and dissolve by heating with frequent agitation. Boil for one minute until complete dissolution. Sterilize in autoclave at 115 °C for 20 minutes. Mix well and dispense into appropriate containers.

Instructions for use

- Consult appropriate references for recommended test procedures.

- Incubate at a temperature of 35±2 °C for 18-24 hours.

Appareance

Quality control

Solubility

Color of the dehvdrated medium

Color of the prepared medium

Final pH (25°C)

w/o rests	Fine powder	Beige	Amber	7,0±0,1
Microbiolo	ogical test			
Incubation con	ditions: (35±2 °C / 18-24	4 h)		
Microorganism	s		Specification	
Escherichia co	li ATCC 23724		Good growth	
Escherichia co	li ATCC 33694		Good growth	
Escherichia co	li ATCC 33849		Good growth	
Escherichia co	li ATCC 39403		Good growth	
Escherichia co	li ATCC 47014		Good growth	

Storage

2YT Autoinducible Growth Medium with Trace Elements

To grow IPTG inducible expression in bacterial strains.

Practical information

Aplications Protein expression Categories Escherichia coli

Industry: Molecular biology / Microbiological Culture Media

Principles and uses

Auto induction media was first formulated and developed by W. Studier to grow IPTG-inducible expression strains. The principle of auto induction media is based on carbon sources in the medium that are metabolized differentially to promote high density cell growth and automatically induce protein expression from lac promoters.

Auto induction media contains glucose as well as lactose as the carbon source. A limited concentration of glucose is metabolized preferentially during growth, which prevents uptake of lactose until the glucose is depleted, usually in mid to late log phase. As the glucose is depleted, lactose can be taken up and converted by the enzyme ß-galactosidase to the inducer allolactose. Allolactose causes the release of lac repressor from its specific binding sites in the DNA and thereby induces expression of T7 RNA polymerase from the lacUV5 promoter and unblocks T7lac promoters, allowing expression of target proteins by T7 RNA polymerase.

With Auto induction media, a high density cell growth is followed by a spontaneous induction of protein expression. There is no need to monitor the cell density and there is no conventional induction with IPTG. Parallel growth of many non-induced or auto-induced cultures is feasible because cultures are simply inoculated and grown to saturation. This is a great convenience and simplifies manual or automated induction and analysis of multiple clones compared to conventional IPTG induction, which requires monitoring growth of each culture and adding inducer at the proper stage of growth.

Formula in g/L

Glucose	0,5	Ammonium sulfate	3,3
Disodium phosphate	7,1	Magnesium sulfate	0,15
Monopotassium phosphate	6,8	Tryptone	16
Yeast extract	10	Trace elements	0,015
Alpha lactose	2		

Preparation

Suspend 45,86 grams of the medium in one liter of distilled water. Mix well and dissolve by heating with frequent agitation. Boil for one minute until complete dissolution. Sterilize in autoclave at 115 °C for 20 minutes. Mix well and dispense into appropriate containers.

Instructions for use

- Consult appropriate references for recommended test procedures.

- Incubate at a temperature of 35±2 °C for 18-48 hours.

Quality control

Solubility	Appareance	Color of the dehydrated medium	Color of the prepared medium	Final pH (25ºC)
w/o rests	Fine powder	Beige	Amber	7,0±0,2

Microbiological test

Incubation conditions: (35±2 °C / 18-48 h)

Mucroorc	ianisms
million oon g	garnorno

Escherichia coli ATCC 23724 Escherichia coli ATCC 33694 Escherichia coli ATCC 33849 Escherichia coli ATCC 39403 Escherichia coli ATCC 47014

Storage

Temp. Min.:2 °C Temp. Max.:25 °C Specification Good growth Good growth Good growth Good growth Good growth

AIM -Super Growth w/o Trace Element

To grow IPTG inducible expression in bacterial strains

Practical information

Aplications

Protein expression

Industry: Culture media for Molecular biology

Categories General use



Principles and uses

Auto Induction Media (AIM) was first formulated and developed by W. studier (1) to grow IPTG-inducible expression strains.

The principle of AIM is based on carbon sources in the medium that are metabolized differentially to promote high density cell growth and automatically induce protein expression from lac promoters. AIM contains glucose as well as lactose as the carbon source. A limited concentration of glucose is metabolized preferentially during growth, which prevents uptake of lactose until the glucose is depleted, usually in mid to late log phase. As the glucose is depleted, lactose can be taken up and converted by the enzyme ß-galactosidase to the inducer allolactose. Allolactose causes the release of lac repressor from its specific binding sites in the DNA and thereby induces expression of T7 RNA polymerase from the lacUV5 promoter and unblocks T7lac promoters, allowing expression of target proteins by T7 RNA polymerase. With AIM media a high density cell growth is followed by a spontaneous induction of protein expression. There is no need to monitor the cell density and there is no conventional induction with IPTG.

The principle of AIM media is based on carbon sources in the medium that are metabolized differentially to promote high density cell growth and automatically induce protein expression from lac promoters. Parallel growth of many non-induced or auto-induced cultures is feasible because cultures are simply inoculated and grown to saturation. This is a great convenience and simplifies manual or automated induction and analysis of multiple clones compared to conventional IPTG induction, which requires monitoring growth of each culture and adding inducer at the proper stage of growth. For higher saturation density than AIM-LB Broth (2094).

Formula in g/L

Glucose	0,5	Alpha-lactose	2
Ammonium sulfate	3,3	Disodium phosphate	7,1
Magnesium sulfate	0,15	Monopotassium phosphate	6,8
Tryptone	35	Yeast extract	20

Preparation

Suspend 74.85 grams of the medium in one liter of distilled water. Mix well and dissolve by heating with frequent agitation. Boil for 1 minute or until complete dissolution. Sterilize in autoclave at 115°C for 20 minutes. Mix well and dispense as wished.

Instructions for use

- Inoculate and incubate at a temperature of 35±2 °C and observed after 18-48 hours.

Quality control

Appareance

Solubility

Color of the dehydrated medium

Color of the prepared medium

Final pH (25°C)

Beige	Amber	7,0 ± 0,1
)		
	Specification	
	Good growth	
-	Beige	Beige Amber Amber Specification Good growth Good grow

Storage

Induction Base Medium

Medium for maintenance and propagation of the PL promoter in E. coli strains GI724, GI826 and GI698.

Practical information

Aplications

Preparation and recovery of competent cells

Categories Escherichia coli

Industry: Molecular biology / Microbiological Culture Media

Principles and uses

Induction Base Medium is used for the maintenance and propagation of the PL promoter in E. coli strains GI724, GI826 and GI698 and to increase plasmid yield during sequencing of positive clones. These strains contain the Lambda cl repressor gene under the control of the tryptophan-inducible trp promoter. This medium has low levels of tryptophan.

Casaminoacids provide the necessary nutrients and cofactors for a good growth of recombinant E. coli strains. Due to their greater degree of digestion, casaminoacids are an excellent source of free amino acids. Phosphates act as a buffer system. Ammonium chloride and magnesium sulfate provide essential ions for transport and osmotic balance.

Formula in g/L

Ammonium chloride	1	Casaminoacids	2
Disodium phosphate	6	Monopotassium phosphate	3
Sodium chloride	0,5	Magnesium chloride	0,095

Preparation

Suspend 12,6 grams of medium in one liter of distilled water. Dissolve by heating with frequent agitation. Boil for one minute until complete dissolution. Sterilize in autoclave at 121 °C for 15 minutes. Cool to room temperature, add 25 ml of a 20% glucose solution and 1 ml of ampicillin (100 g/ml) under sterile conditions. Mix well.

Instructions for use

Consult an appropriate reference for recommended test procedures.

Quality control

Solubility	Appareance	Color of the dehydrated medium	Color of the prepared medium	Final pH (25°C)
w/o rests	Fine powder	Beige	Amber	7,0±0,2

Microbiological test

Incubation conditions: (35±2 °C / 18-24 h).		
Microorganisms	Specification	
Escherichia coli ATCC GI724	Good growth	

Storage

LB Agar (Lennox)

Recommended medium for maintaining and cultivating recombinant strains of E. coli.

Practical information

Aplications Categories Preparation and recovery of competent cells Escherichia coli Industry: Culture media for Molecular biology

Principles and uses

LB Agar (Lennox) is a nutritionally rich medium developed by Lennox for the growth and maintenance of pure cultures of recombinant strains of E. coli used inused in molecular microbiology procedures.

These strains are generally derived from E. coli K12, which are unable to produce vitamin B, so this media is formulated to enhance the growth of nutritionally demanding microorganisms. This strain of E. coli has been further modified through specific mutation to create an auxotrophic strain that is not capable of growth on nutritionally deficient media.

Tryptone provides nitrogen, vitamins, minerals and amino acids essential for growth. Yeast extract is source of vitamins, particularly the B-group. Sodium chloride supplies essential electrolytes for transport and osmotic balance. Bacteriological agar is the solidifying agent. If desired, antibiotics can also be added.

LB Agar (Lennox) has a different sodium chloride level than other media such as Luria Agar (Miller LB Agar) (Cat. 1552) or Luria Agar (Miller Modification) (Cat. 1308). This allows to select the optimum salt concentration of the medium for a specific strain.

Formula in g/L

Bacteriological agar	15	Sodium chloride	5
Tryptone	10	Yeast extract	5

Preparation

Suspend 35 grams of medium in one liter of distilled water. Mix well and dissolve by heating with frequent agitation. Boil for one minute until complete dissolution. Sterilize in autoclave at 121 °C for 15 minutes. Cool to 45-50 °C, mix well and dispense into plates.

Instructions for use

Carry out the experimental procedure according to appropriate use or purpose.
 Inoculate and incubate at a temperature of 35±2 °C for 18-24 hours.

Quality control

Solubility	Appareance	Color of the dehydrated medium	Color of the prepared medium	Final pH (25°C)
w/o rests	Fine powder	Beige	Amber, slightly opalescent	7,0±0,2



Microbiological test

Incubation conditions: (35±2 °C / 18-24 h).

Microorganisms	Specification
Escherichia coli ATCC 23724	Good growth
Escherichia coli ATCC 33694	Good growth
Escherichia coli ATCC 33849	Good growth
Escherichia coli ATCC 39403	Good growth
Escherichia coli ATCC 47014	Good growth

Storage

LB Agar (Lennox) with Ampicilin 100 µg/ml

For the cultivation of E. coli in molecular genetics studies.

Practical information

Aplications

Selection of transformants

Categories Escherichia coli

Industry: Molecular biology / Microbiological Culture Media

Principles and uses

The LB Agar (Lennox) with Ampicilin 100 µg/ml medium is used for the selective growth of ampicillin resistant E. coli recombinant strains in molecular genetic studies. This medium is recommended for strains that require less salt concentration.

The transformed E. coli are plated directly onto selective agar media (LB Agar containing antibiotic), fewer transformed colonies will appear per ml plated. To select the bacteria with the plasmid, it is necessary to subcultivate an inoculum from LB agar to a LB broth with the antibiotic added.

Formula in g/L

Ampicillin	0,1	Bacteriological agar	15
Sodium chloride	5	Tryptone	10
Yeast extract	5		

Preparation

Suspend 35 grams of medium in one liter of distilled water. Mix well and dissolve by heating with frequent agitation. Boil for one minute until complete dissolution. AVOID OVERHEATING. DO NOT AUTOCLAVE. Cool to 45-50 °C, mix well and dispense into plates.

Instructions for use

Inoculate and incubate at a temperature of 35±2 °C for 18-24 hours.

Quality control

Solubility	Appareance	Color of the dehydrated medium	Color of the prepared medium	Final pH (25ºC)
w/o rests	Fine powder	Beige	Amber, slightly opalescent	

Microbiological test

Incubation conditions: (35±2 °C / 18-24 h).

Microorganisms	Specification
Escherichia coli DH5 alpha + pUC19	Good growth
Escherichia coli ATCC 25922	Total inhibition
Escherichia coli ATCC 8739	Total inhibition

<u>Storage</u>

Temp. Min.:2 °C Temp. Max.:8 °C

LB Agar with Kanamycin 50 µg/ml (Lennox)

To select colonies of Escherichia coli in molecular genetic studies.

Practical information

Aplications

Selection of transformants

Categories Escherichia coli

Industry: Molecular biology / Microbiological Culture Media

Principles and uses

LB Agar with Kanamycin 50 µg/ml (Lennox) is used for the selective growth of Kanamycin resistant E. coli recombinant strains in molecular genetic studies. This medium is recommended for strains that require less salt concentration.

The transformed E. coli are plated directly onto selective agar media (LB Agar containing antibiotic), where fewer transformed colonies will appear per ml plated. To select the bacteria with the plasmid, it is necessary to subcultivate an inoculum from LB agar to a LB broth with the antibiotic added.

Formula in g/L

Bacteriological agar	15	Kanamycin	0,05
Sodium chloride	5	Tryptone	10
Yeast extract	5		

Preparation

Suspend 35 grams of medium in one liter of distilled water. Mix well and dissolve by heating with frequent agitation. Boil for one minute until complete dissolution. Sterilize in autoclave at 118 °C for 10 minutes. Cool to 45-50 °C, mix well and dispense into plates.

Instructions for use

- Carry out the experimental procedure according to appropriate use or purpose.

- Inoculate and incubate at a temperature of 35±2 °C for 18-24 hours.

Quality control

Solubility	Appareance	Color of the dehydrated medium	Color of the prepared medium	Final pH (25°C)
w/o rests	Fine powder	Beige	Amber, slightly opalescent	7,0±0,2

Microbiological test

Incubation conditions: (35±2 °C / 18-24 h).

Microorganisms	Specification	
Escherichia coli DH5 alpha + PH SG 298	Good growth	
Escherichia coli ATCC 25922	Total inhibition	
Escherichia coli ATCC 8739	Total inhibition	

<u>Storage</u>

LB Broth (Lennox)

Recommended medium for maintaining and cultivating recombinant strains of E. coli.

Practical information

Aplications Preparation and recovery of competent cells Categories Escherichia coli

Industry: Microbiological Culture Media

Principles and uses

LB Broth (Lennox) is a nutritionally rich medium developed by Lennox for the growth and maintenance of pure cultures of recombinant strains of E. coli used in molecular and microbiological procedures.

These strains are generally derived from E. coli K12, which are unable to produce vitamin B, so this media is formulated to enhance the growth of nutritionally demanding microorganisms. This strain of E. coli has been further modified through specific mutation to create an auxotrophic strain that is not capable of growth on nutritionally deficient media. Cultivation in LB Broth allows cells with an insert plasmid to start expressing the genes on the transformed plasmid, including the antibiotic resistance gene. If transformed E. coli are plated directly onto selective agar media (LB Agar containing antibiotic), fewer transformed colonies will appear per ml plated. Growing the transformed cells in LB broth will increase the number of transformed cells per ml.

LB Broth (Lennox) contains ten times the sodium chloride level of Luria Broth (Miller's Modification) (Cat. 1266) and a half of the level found in Luria Broth (Miller's LB Broth) (Cat. 1551). This allows selecting the optimal salt concentration medium for a specific strain.

Tryptone provides nitrogen, vitamins, minerals and amino acids essential for growth. Yeast extract is source of vitamins, particularly the B-group. Sodium chloride supplies essential electrolytes for transport and osmotic balance. This medium consist of the same ingredients as LB Agar (Lennox) without bacteriological agar. If desired, antibiotics can also be added.

Formula in g/L

Sodium chloride	5	Tryptone	10
Yeast extract	5	_	

Preparation

Suspend 20 grams of the medium in one liter of distilled water. Mix well and dissolve by heating with frequent agitation. Boil for one minute until complete dissolution. Dispense into appropriate containers and sterilize in autoclave at 121 °C for 15 minutes.

Instructions for use

- Carry out the experimental procedure according to appropriate use or purpose.

- Inoculate and incubate at a temperature of 35±2 °C for 18-24 hours.

Quality control

Solubility	Appareance	Color of the dehydrated medium	Color of the prepared medium	Final pH (25ºC)



w/o rests	Fine powder	Beige	Clear amber	7,0±0,2
Microbiol	ogical test			
Incubation cor	nditions: (35±2 °C / 18-2	4 h).		
Microorganisn	ns		Specification	
Escherichia co	oli ATCC 23724		Good growth	
Escherichia co	oli ATCC 33694		Good growth	
Escherichia co	oli ATCC 33849		Good growth	
Escherichia co	oli ATCC 39403		Good growth	
Escherichia co	bli ATCC 47014		Good growth	
Storage				

LB Broth Autoinducible w/o Trace Element

For autoinducible expression of IPTG-inducible bacterial strains

Practical information

Aplications

Protein expression

Categories Escherichia coli

Industry: Molecular biology / Microbiological Culture Media

Principles and uses

LB Broth Autoinducible w/o Trace Element is a medium which supports a high cell density and, in this case, it is formulated for the optimum growth of E.coli during the logarithmic phase for a long time. As a result, it yields a greater number of recombinant proteins and plasmic DNA.

Auto induction media was first formulated and developed by W. studier to grow IPTG-inducible expression strains. The principle of auto induction media is based on carbon sources in the medium that are metabolized differentially to promote high density cell growth and automatically induce protein expression from lac promoters. Auto induction media contains glucose as well as lactose as the carbon source. A limited concentration of glucose is metabolized preferentially during growth, which prevents uptake of lactose until the glucose is depleted, usually in mid to late log phase. As the glucose is depleted, lactose can be taken up and converted by the enzyme ß-galactosidase to the inducer allolactose. Allolactose causes the release of lac repressor from its specific binding sites in the DNA and thereby induces expression of T7 RNA polymerase from the lacUV5 promoter and unblocks T7lac promoters, allowing expression. There is no need to monitor the cell density and there is no conventional induction with IPTG.

Parallel growth of many non-induced or auto-induced cultures is feasible because cultures are simply inoculated and grown to saturation. This is a great convenience and simplifies manual or automated induction and analysis of multiple clones compared to conventional IPTG induction, which requires monitoring growth of each culture and adding inducer at the proper stage of growth.

Formula in g/L

Glucose	0,5	Alpha-lactose	2
Ammonium sulfate	3,3	Disodium phosphate	7,1
Magnesium sulfate	0,15	Monopotassium phosphate	6,8
Tryptone	10	Yeast extract	5

Preparation

Suspend 34,85 grams of the medium in one liter of distilled water. Mix well and dissolve by heating with frequent agitation. Boil for 1 minute or until complete dissolution. Sterilize in autoclave at 121 °C for 15 minutes. Mix well and dispense as wished.

Instructions for use

- Carry out the experimental procedure according to appropriate use or purpose.

- Inoculate and incubate at a temperature of 35±2 °C for 18-48 hours.

Quality control

Solubility	Appareance	Color of the dehydrated medium	Color of the prepared medium	Final pH (25°C)
w/o rests	Fine powder	Beige	Amber	7,0±0,2

Microbiological test

Incubation conditions: (35±2 °C / 18-48 h).

Mucroorc	ianisms
million oon g	garnorno

Escherichia coli ATCC 23724 Escherichia coli ATCC 33694 Escherichia coli ATCC 33849 Escherichia coli ATCC 39403 Escherichia coli ATCC 47014

Storage

Temp. Min.:2 °C Temp. Max.:25 °C Specification Good growth Good growth Good growth Good growth Good growth

LB Broth Autoinducible with Trace Element

For expression of IPTG-inducible bacterial strains.

Practical information

Aplications Protein expression Categories Escherichia coli

Industry: Culture media for Molecular biology



Principles and uses

LB Broth Autoinducible with Trace Element is a medium which supports a high cell density and, in the case is formulated for the optimum growth of E.coli, maintains growth in the logarithmic phase for a long time. As a result, it yields a greater number of recombinant proteins and plasmic DNA.

Autoinduction Media (AIM) was first formulated and developed by W. studier to grow IPTG-inducible expression strains. The principle of AIM is based on carbon sources in the medium that are metabolized differentially to promote high density cell growth and automatically induce protein expression from lac promoters. AIM contains glucose as well as lactose as the carbon source. A limited concentration of glucose is metabolized preferentially during growth, which prevents uptake of lactose until the glucose is depleted, usually in mid to late log phase. As the glucose is depleted, lactose can be taken up and converted by the enzyme &-galactosidase to the inducer allolactose. Allolactose causes the release of lac repressor from its specific binding sites in the DNA and thereby induces expression of T7 RNA polymerase from the lacUV5 promoter and unblocks T7Iac promoters, allowing expression. There is no need to monitor the cell density and there is no conventional induction with IPTG. Parallel growth of many non-induced or auto-induced cultures is feasible because cultures are simply inoculated and grown to saturation. This is a great convenience and simplifies manual or automated induction and analysis of multiple clones compared to conventional IPTG induction, which requires monitoring growth of each culture and adding inducer at the proper stage of growth.

Tryptone provide nitrogen, vitamins, minerals and amino acids essential for growth. Yeast extract is source of vitamins, particularly the B-group. Potassium phosphates act as a buffer system to prevent cell death. Its formulation is characterized by the presence of trace elements which supply all of the specific requirements for the bacteria.

Formula in g/L

Glucose 0,5	Ammonium sulfate 3,3
Disodium phosphate 7,1	Magnesium sulfate 0,15
Monopotassium phosphate 6,8	Tryptone 10
Yeast extract 5	Trace elements 0,015
Alpha lactose 2	_

Preparation

Suspend 34,9 grams of medium in one liter of distilled water. Mix well and dissolve by heating with frequent agitation. Boil for one minute until complete dissolution. Sterilize in autoclave at 121 °C for 15 minutes. Mix well and dispense as wished.

Instructions for use

- Carry out the experimental procedure according to appropriate use or purpose.
- Inoculate and incubate at a temperature of 35±2 °C for 18-48 hours.

Quality of	control
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Solubility	Appareance	Color of the dehydrated medium	Color of the prepared medium	Final pH (25°C)
w/o rests	Fine powder	Beige	Amber	7,0±0,2

Microbiological test

Incubation conditions: (35±2 °C / 18-48 h).

Microorganisms	Specification
Escherichia coli ATCC 23724	Good growth
Escherichia coli ATCC 33694	Good growth
Escherichia coli ATCC 33849	Good growth
Escherichia coli ATCC 39403	Good growth
Escherichia coli ATCC 47014	Good growth

Storage

Luria Agar LB Agar)

Recommended medium for maintaining and cultivating recombinant strains of E. coli.

Practical information

Aplications

Preparation and recovery of competent cells

Categories Escherichia coli

Industry: Culture media for Molecular biology

Principles and uses

Luria Agar (Miller's LB Agar) is based on LB Medium as described by Miller for the growth and maintenance of E. coli strains used in molecular microbiology procedures.

These strains are generally derived from E. coli K12, which are unable to produce vitamin B, so this media is formulated to enhance the growth of nutritionally demanding microorganisms. This strain of E. coli has been further modified through specific mutation to create an auxotrophic strain that is not capable of growth on nutritionally deficient media.

Tryptone provides nitrogen, vitamins, minerals and amino acids essential for growth. Yeast extract is source of vitamins, particularly the B-group. Sodium chloride supplies essential electrolytes for transport and osmotic balance. Bacteriological agar is the solidifying agent.

If desired aseptically add 10 ml of sterile 20% glucose solution and mix thoroughly for a better growth. Bacteria that contain plasmids tend to grow best in broth that has between 5 and 10 g of salt. Various cofactors may also need to be added to the broth if working with certain types of bacteriophages. For example, bacteriophage labmda requires an excess of magnesium in the broth to properly infect bacteria.

Luria Agar (Miller LB Agar) has a different sodium chloride level than other media such as LB Agar (Lennox) (Cat. 1083) or Luria Agar (Miller Modification) (Cat. 1308). This allows to select the optimum salt concentration of the medium for a specific strain.

Formula in g/L

Bacteriological agar 15	Sodium chloride	10
Tryptone 10	Yeast extract	5

Typical formula g/L * Adjusted and/or supplemented as required to meet performance criteria.

Preparation

Suspend 40 grams of medium in one liter of distilled water. Mix well and dissolve by heating with frequent agitation. Boil for one minute until complete dissolution. Sterilize in autoclave at 121 °C for 15 minutes. Cool to 45-50 °C, mix well and dispense into plates.

Instructions for use

- Carry out the experimental procedure according to appropriate use or purpose.

- Inoculate and incubate at a temperature of 35±2 °C for 18-24 hours.

Quality control

Solubility	Appareance	Color of the dehydrated medium	Color of the prepared medium	Final pH (25°C)
w/o rests	Fine powder	Beige	Amber, slightly opalescent	7,0±0,2

Microbiological test

Incubation conditions: $(35\pm 2 \text{ °C} / 18-24 \text{ h})$ Inoculation conditions: Productivity quantitative (100±20. Min.50 cfu)

Reference medium: TSA

Microorganisms

Escherichia coli ATCC 23724 Escherichia coli ATCC 33694 Escherichia coli ATCC 33849 Escherichia coli ATCC 39403 Escherichia coli ATCC 47014

Storage

Temp. Min.:2 °C Temp. Max.:25 °C

Specification

Good growth >70% Good growth >70% Good growth >70% Good growth >70% Good growth >70%

Luria Agar Modification)

Recommended medium for maintaining and cultivating recombinant strains of E. coli.

Practical information

Aplications

Preparation and recovery of competent cells

Categories Escherichia coli

Industry: Microbiological Culture Media

Principles and uses

Luria Agar (Miller's Modification) is based on LB Medium according to Miller's descrption. Its modification consists of a minimal concentration of sodium chloride. This medium is used for the growth and maintenance of E. coli strains used in molecular microbiology procedures. It is used for strains in which the optimal concentration of salt is 0,5 g/l.

These strains are generally derived from E. coli K12, which are unable to produce vitamin B, so this media is formulated to enhance the growth of nutritionally demanding microorganisms. This strain of E. coli has been further modified through specific mutation to create an auxotrophic strain that is not capable of growth on nutritionally deficient media. Some plasmid vectors replicate to high copy numbers and do not require selective amplification. Some vectors do not replicate so freely and need to be selectively amplified. Antibiotics may be added to inhibit host synthesis and, as a result, prevent replication of the bacterial chromosomome.

Tryptone provides nitrogen, vitamins, minerals and amino acids essential for growth. Yeast extract is source of vitamins, particularly the B-group. Sodium chloride supplies essential electrolytes for transport and osmotic balance. Bacteriological agar is the solidifying agent.

Luria Agar (Miller's Modification) has a different sodium chloride level than other media such as LB Agar (Lennox) (Cat. 1083) or Luria Agar (Miller's LB Agar) (Cat. 1552). This allows to select the optimum salt concentration of the medium for a specific strain.

Formula in g/L

Bacteriological agar	15	Sodium chloride	0,5
Tryptone	10	Yeast extract	5

Preparation

Suspend 30,5 grams of medium in one liter of distilled water. Mix well and dissolve by heating with frequent agitation. Boil for one minute until complete dissolution. Sterilize in autoclave at 121 °C for 15 minutes. Cool to 45-50 °C, mix well and dispense into plates.

Instructions for use

- Carry out the experimental procedure according to appropriate use or purpose.

- Inoculate and incubate at a temperature of 35±2 °C for 18-24 hours.

Quality control

Solubility	Appareance	Color of the dehydrated medium	Color of the prepared medium	Final pH (25°C)
w/o rests	Fine powder	Beige	Amber, slightly opalescent	7,0±0,2

Microbiological test

Incubation conditions: (35±2 °C / 18-24 h)

Microorganisms

Specification

Escherichia coli ATCC 23724 Escherichia coli ATCC 33694 Escherichia coli ATCC 33849 Escherichia coli ATCC 39403 Escherichia coli ATCC 47014

Storage

Temp. Min.:2 °C Temp. Max.:25 °C Good growth Good growth Good growth Good growth Good growth

Luria Agar with Ampicillin 100 μ g/ml (Miller's LB Agar)

For molecular studies of E. coli.

Practical information

Aplications

Selection of transformants

Categories Escherichia coli

Industry: Culture media for Molecular biology

Principles and uses

Luria Agar with Ampicillin 100 µg/ml (Miller's LB Agar) medium is used for the selective growth of ampicillin resistant E. coli recombinant strains in molecular genetic studies.

The transformed E. coli are plated directly onto selective agar media (LB Agar containing antibiotic), fewer transformed colonies will appear per ml plated. To select the bacteria with the plasmid, it is necessary to subcultivate an inoculum from LB Agar to a LB Broth with the antibiotic added.

Formula in g/L

Ampicillin 0,1	Bacteriological agar 15
Sodium chloride 10	Tryptone 10
Yeast extract 5	

Typical formula g/L * Adjusted and/or supplemented as required to meet performance criteria.

Preparation

Suspend 40 grams of medium in one liter of distilled water. Mix well and dissolve by heating with frequent agitation. Boil for one minute until complete dissolution. DO NOT OVERHEAT. DO NOT AUTOCLAVE. Cool to 45-50 °C, mix well and dispense into plates.

Instructions for use

- Carry out the experimental procedure according to apprpriate use or purpose.

- Inoculate and incubate at 35±2 °C for 18-24 hours.

Quality control

Solubility	Appareance	Color of the dehydrated medium	Color of the prepared medium	Final pH (25°C)
w/o rests	Fine powder	Beige	Amber, slightly opalescent	7,0±0,2

Microbiological test

Incubation conditions: (35±2 °C / 18-24 h) Inoculation conditions: Productivity quantitative (100±20. Min.50 cfu) / Selectivity (10^4-10^6 cfu) Reference medium: TSA

Microorganisms	Specification
Escherichia coli DH5 alpha + pUC19	Good growth >50%
Escherichia coli ATCC 25922	Total inhibition
Escherichia coli ATCC 8739	Total inhibition

Storage

Luria Agar with Ampicillin 50 µg/ml (Miller's LB Agar)

For E.coli in molecular genetics studies.

Practical information

Aplications

Selection of transformants

Industry: Culture media for Molecular biology

Principles and uses

Luria Agar with Ampicillin 50 µg/ml (Miller's LB Agar) is used for the selective growth of ampicillin resistant E. coli recombinant strains in molecular genetic studies.

The transformed E. coli are plated directly onto selective agar media (LB Agar containing antibiotic), fewer transformed colonies will appear per ml plated. To select the bacteria with the plasmid, it is necessary to subcultivate an inoculum from LB Agar to a LB Broth with the antibiotic added.

Tryptone provide nitrogen, vitamins, minerals and amino acids essential for growth. Yeast extract is source of vitamins, particularly the B-group. Sodium chloride provides osmotic balance. Bacteriological agar is the solidifying agent.

Formula in g/L

Ampicillin	0,05	Bacteriological agar	15
Sodium chloride	10	Tryptone	10
Yeast extract	5		

Typical formula g/L * Adjusted and/or supplemented as required to meet performance criteria.

Preparation

Suspend 40 grams of medium in one liter of distilled water. Mix well and dissolve by heating with frequent agitation. Boil for one minute until complete dissolution. Do not overheat. Do not sterilize in autoclave. Cool to 45-50 °C, mix well and dispense into plates.

Instructions for use

- Carry out the experimental procedure according to apprpriate use or purpose.

- Inoculate and incubate at 35±2 °C for 18-24 hours.

Quality control

Solubility	Appareance	Color of the dehydrated medium	Color of the prepared medium	Final pH (25⁰C)
w/o rests	Fine powder	Beige	Ambre, slightly opalescent	7,0±0,2

Microbiological test

Incubation conditions: (35±2 °C / 18-24 h) Inoculation conditions: Productivity quantitative (100±20. Min.50 cfu) / Selectivity (10^4-10^6 cfu) Reference medium: TSA

Microorganisms Escherichia coli DH5 alpha + pUC19 Escherichia coli ATCC 25922

Good growth >50%
Total inhibition

Specification

Categories Escherichia coli

Storage

Luria Agar with Kanamycin 50 µg/ml (Miller's LB Agar)

To select colonies of Escherichia coli in molecular genetics.

Practical information

Industry: Culture media for Molecular biology

Aplications Selection of transformants Categories Escherichia coli



Principles and uses

Luria Agar with Kanamycin 50 µg/ml (Miller's LB Agar) is used for the selective growth of Kanamycin resistant E. coli recombinant strains in molecular genetic studies. This medium is recommended for strains that require less salt concentration.

The transformed E. coli are plated directly onto selective agar media (LB Agar containing antibiotic), where fewer transformed colonies will appear per ml plated. To select the bacteria with the plasmid, it is necessary to subcultivate an inoculum from LB Agar to LB Broth with the antibiotic added.

Formula in g/L

Bacteriological agar	15	Kanamycin	0,05
Sodium chloride	10	Tryptone	10
Yeast extract	5		

Typical formula g/L * Adjusted and/or supplemented as required to meet performance criteria.

Preparation

Suspend 40 grams of medium in one liter of distilled water. Mix well and dissolve by heating with frequent agitation. Boil for one minute until complete dissolution. DO NOT OVERHEAT. DO NOT AUTOCLAVE. Cool to 45-50 °C, mix well and dispense into plates.

Instructions for use

- Carry out the experimental procedure according to apprpriate use or purpose.

- Inoculate and incubate at 35±2 °C for 18-24 hours.

Quality control

Solubility	Appareance	Color of the dehydrated medium	Color of the prepared medium	Final pH (25°C)
w/o rests	Fine powder	Beige	Amber, slightly opalescent	7,0±0,2

Microbiological test

Incubation conditions: (35±2 °C / 18-24 h) Inoculation conditions: Productivity quantitative (100±20. Min.50 cfu) / Selectivity (10^4-10^6 cfu)

Microorganisms

Escherichia coli DH5 alpha + PH SG 298 Escherichia coli ATCC 25922 Escherichia coli ATCC 8739

Storage

Temp. Min.:2 °C Temp. Max.:8 °C Specification

Good growth >50% Total inhibition Total inhibition

Luria Broth (Miller's LB Broth)

For molecular genetics studies in E.coli

Practical information

Principles and uses

microbiology procedures.

Aplications

Prenaration and	recovery of competent cells	

Industry: Culture media for Molecular biology

Categories Escherichia coli



If desired aseptically add 10 ml of sterile 20% glucose solution and mix thoroughly for a better growth. Bacteria that contain plasmids tend to grow best in broth that has between 5 and 10 g of salt. Various cofactors may also need to be added to the broth if working with certain types of bacteriophages. For example, bacteriophage labmda requires an excess of magnesium in the broth to properly infect bacteria.

Luria Broth (Miller's LB Broth) is based on LB Medium as described by Miller for the growth and maintenance of E. coli strains used in molecular

These strains are generally derived from E. coli K12, which are unable to produce vitamin B, so this media is formulated to enhance the growth of

Luria Broth (Miller's LB Broth) has a different sodium chloride level than other media such as LB Broth (Lennox) (Cat. 1231) or Luria Broth (Miller's Modification) (Cat. 1266). This allows to select the optimum salt concentration of the medium for a specific strain.

Formula in g/L

Sodium chloride	10 Tryptone	10
Yeast extract	5	

Typical formula g/L * Adjusted and/or supplemented as required to meet performance criteria.

Preparation

Suspend 25 grams of the medium in one liter of distilled water. Mix well and dissolve by heating with frequent agitation. Boil for one minute until complete dissolution. Dispense into appropriate containers and sterilize in autoclave at 121 °C for 15 minutes.

Instructions for use

- Carry out the experimental procedure according to appropriate use or purpose.

- Inoculate and incubate at a temperature of 35±2 °C for 18-24 hours.

Quality control

Solubility	Appareance	Color of the dehydrated medium	Color of the prepared medium	Final pH (25⁰C)
w/o rests	Fine powder	Beige	Amber	7,0±0,2

Microbiological test

Incubation conditions: (35±2 °C / 18-24 h) Inoculation conditions: Productivity qualitative (<100 cfu)

Microorganisms	Specification
Escherichia coli ATCC 23724	Good growth /Turbidity (2)
Escherichia coli ATCC 33694	Good growth / Turbidity (2)
Escherichia coli ATCC 33849	Good growth / Turbidity (2)
Escherichia coli ATCC 39403	Good growth / Turbidity (2)
Escherichia coli ATCC 47014	Good growth / Turbidity (2)

Storage

Luria Broth Modification)

Recommended medium for maintaining and cultivating recombinant strains of E. coli.

Practical information

Aplications Preparation and recovery of competent cells Categories Escherichia coli

Industry: Culture media for Molecular biology

Principles and uses

Luria Broth (Miller's Modification) is based on LB Medium according to Miller's descrption. Its modification consists of a minimal concentration of sodium chloride. This medium is used for the growth and maintenance of E. coli strains used in molecular microbiology procedures. It is used for strains in which the optimal concentration of salt is 0,5 g/l.

These strains are generally derived from E. coli K12, which are unable to produce vitamin B, so this media is formulated to enhance the growth of nutritionally demanding microorganisms. This strain of E. coli has been further modified through specific mutation to create an auxotrophic strain that is not capable of growth on nutritionally deficient media. Some plasmid vectors replicate to high copy numbers and do not require selective amplification. Some vectors do not replicate so freely and need to be selectively amplified. Antibiotics may be added to inhibit host synthesis and, as a result, prevent replication of the bacterial chromosomome.

Tryptone provides nitrogen, vitamins, minerals and amino acids essential for growth. Yeast extract is source of vitamins, particularly the B-group. Sodium chloride supplies essential electrolytes for transport and osmotic balance.

Luria Broth (Miller's Modification) has a different sodium chloride level than other media such as LB Broth (Lennox) (Cat. 1231) or Luria Broth (Miller's LB Broth) (Cat. 1551). This allows to select the optimum salt concentration of the medium for a specific strain.

Formula in g/L

Sodium chloride 0	5 Tryptone 10
Yeast extract	5

Preparation

Suspend 15,5 grams of the medium in one litre of distilled water. Mix well and dissolve by heating with frequent agitation. Boil for one minute until complete dissolution. Dispense into appropriate containers and sterilize in autoclave at 121 °C for 15 minutes.

Instructions for use

- Carry out the experimental procedure according to appropriate use or purpose.

- Inoculate and incubate at a temperature of 35±2 °C for 18-24 hours.

Quality control

Solubility	Appareance	Color of the dehydrated medium	Color of the prepared medium	Final pH (25°C)

w/o rests	Fine powder	Beige	Amber, slightly opalescent	7,0±0,2
Microbiol	ogical test			
Incubation cor	nditions: (35±2 °C / 18-2	4 h).		
Microorganism	ns		Specification	
Escherichia co	oli ATCC 23724		Good growth	
Escherichia co	oli ATCC 33694		Good growth	
Escherichia co	oli ATCC 33849		Good growth	
Escherichia co	oli ATCC 39403		Good growth	
Escherichia co	oli ATCC 47014		Good growth	

Storage
Modified Terrific Broth

Medium for use with glycerol in the culture of recombinant E.coli strains.

Practical information

Aplications Preparation and recovery of competent cells

Industry: Culture media for Molecular biology

Categories Escherichia coli





Principles and uses

Modified Terrific Broth is a medium that supports a high cell density and, when formulated for optimum growth of E.coli, maintains growth in the logarithmic phase for a long time. As a result, it yields a greater number of recombinant proteins and plasmic DNA. In some circumstances it substitutes LB Broth (Lennox) (Cat. 1231) used in genetic studies.

Tryptone provide nitrogen, vitamins, minerals and amino acids essential for growth Yeast extract is source of vitamins, particularly the B-group. Potassium phosphates act as a buffer system to prevent cell death.

Glycerol is the source of carbohydrates and carbon since it is not fermented to acetic acid as glucose and does not lead to confusing results.

The formulation of this medium was modified from the Terrific Broth (Cat. 1246) with a different concentration of biological buffer.

Formula in g/L

Dipotassium phosphate	9,4	Monopotassium phosphate	2,2
Tryptone	12	Yeast extract	24

Preparation

Suspend 47.6 grams of the medium in 900 ml of distilled water .Mix well. Add 4 ml of glycerol, adjust to a final volume of 1000 ml, and dissolve by heating with frequent agitation. Boil for one minute until complete dissolution. Dispense into appropriate containers and sterilize in autoclave at 121°C for 15 minutes.

Instructions for use

- Carry out the experimental procedure according to appropriate use or purpose.

- Inoculate and incubate at a temperature of 35±2 °C for 18-24 hours.

Quality control

Solubility	Appareance	Color of the dehydrated medium	Color of the prepared medium	Final pH (25⁰C)
w/o rests	Fine powder	Beige	Amber	7,2 ± 0,2

Microbiological test

Incubation conditions: (35±2 °C / 18-24 h)

Microorganisms

Escherichia coli ATCC 23724 Escherichia coli ATCC 33694 Escherichia coli ATCC 33849 Escherichia coli ATCC 39403 Escherichia coli ATCC 47014

Storage

Temp. Min.:2 °C Temp. Max.:25 °C Specification Good growth Good growth Good growth Good growth Good growth

NZCYM Broth

For the cultivation of recombinant E.coli strains.

Practical information

Aplications Preparation and recovery of competent cells

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Industry: Culture media for Molecular biology

Escherichia coli

Categories



Principles and uses

NZCYM Broth is used as an enrichment medium for the cultivation of recombinant strains of E.coli and propagating lambda bacteriophages developed by Blattner and colleagues.

E.coli grows fast in this enrichment medium, which supplies the amino acids, vitamins and other metabolites as nucleotide precursors and other factors that otherwise would be synthesized by the cell. Casein digest, Yeast extract and Casamino acids provide the necessary nutrients and cofactors required for excellent growth of recombinant strains of E.coli. Magnesium sulfate is the magnesium ions font required in a big variation of enzymatic reactions, including DNA replication.

Formula in g/L

Casaminoacids	1	Magnesium sulfate	0,98
Pancreatic digest of casein	10	Sodium chloride	5
Yeast extract	5		

Preparation

Suspend 22 grams of the medium in one liter of distilled water. Mix well and dissolve by heating with frequent agitation. Boil for one minute until complete dissolution. Dispense into appropriate containers and sterilize in autoclave at 121°C for 15 minutes.

Instructions for use

- Carry out the experimental procedure according to appropriate use or purpose.

- Inoculate and incubate at a temperature of 35±2 °C for 18-24 hours.

Quality control

Solubility	Appareance	Color of the dehydrated medium	Color of the prepared medium	Final pH (25°C)
w/o rests	Fine powder	Beige	Amber	7,0 ± 0,2

Microbiological test

Incubation conditions: (35±2 °C / 18-24 h)

Mucroorc	ianisms
million oon g	garnorno

Escherichia coli ATCC 23724 Escherichia coli ATCC 33694 Escherichia coli ATCC 33849 Escherichia coli ATCC 39403 Escherichia coli ATCC 47014

Storage

Temp. Min.:2 °C Temp. Max.:25 °C Specification Good growth Good growth Good growth Good growth Good growth

RM Base Medium

Solid medium for the maintenance and propagation of the promoter PL in the E. coli strains GI724, GI826 and GI698

Practical information

Aplications

Preparation and recovery of competent cells

Industry: Microbiological Culture Media

Principles and uses

RM Base Medium is used for the maintenance and propagation of the promoter PL in E. coli strains GI724, GI826 and GI698 and to increase the yield of plasmid for sequencing positive clones. These strains have the gene Lambda cl repressor, under the control of the promoter tryptophane inducible, trp. This medium has low tryptophane levels.

Casaminoacids provides the necessary nutrients and cofactors required for excellent growth of recombinant strains of E. coli. Due to its higher degree of digestion, casaminoacids are an excellent source of free aminoacids. Phosphates act as a buffer system. Ammonium chloride and magnesium sulfate provide essential ions. Sodium chloride supplies essential electrolytes for transport and osmotic balance. To promote growth it may require the addition of glucose.

Formula in g/L

Ammonium chloride	1	Casaminoacids	20
Disodium phosphate	6	Magnesium citrate	0,095
Sodium chloride	0,5	Monosodium Phosphate	3

Preparation

Suspend 30.6 grams of the dehydrated medium in 900 ml of distilled water, add 20 ml of 50% glycerol and adjust to a final volume of 1000 ml. Mix well. Heat with frequent agitation until complete dissolution. Distribute in appropriate containers and sterilize in the autoclave at 121 °C for 15 minutes. Add 1 ml/liter of 100 µg/ml of ampicillin under sterile conditions and mix well.

Instructions for use

- Inoculate and incubate at a temperature of 35±2 °C for 18 -24 hours.

Quality control

Solubility	Appareance	Color of the dehydrated medium	Color of the prepared medium	Final pH (25°C)
w/o rests	Fine powder	Beige	Amber	7,0 ± 0,2

Microbiological test

Incubation conditions: (35±2 °C / 18-24 h).	
Microorganisms	Specification
Escherichia coli ATCC GI724	Good growth

Storage

Temp. Min.:2 °C Temp. Max.:25 °C

Cat. 1538

Categories Escherichia coli

sLB Agar

Medium designed to increase bacterial growth and leads to high yield of low copy plasmids and extra high yields of high copy plasmids.

Practical information

Aplications

Preparation and recovery of competent cells

Categories Escherichia coli

Industry: Molecular biology / Microbiological Culture Media

Principles and uses

sLB Agar has been formulated to significantly increase cellular density when compared to the traditional LB Agar. In the standard LB Agar, E. coli cells reach an abrupt stationary phase upon consumption of nutrients contained in the medium. Cell multiplication is stopped, some cell die and plasmid are lost.

Based on the findings of extensive research, our laboratories have developed a new formulation using a proprietary peptone mixture, yeast extract and salts which allow recombinant E. coli cells to have a higher growth. At the end of the log phase replication continues, thus obtaining higher DNA plasmid yields.

sLB Agar cultures have shown cell stability up to 3 days without cell death, being this one a more convenient medium that eliminates the need of constant attention. E.coli's growth is higher in sLB mediums than in standard LB after 3 days at 37 °C.

The special peptone mixture, yeast extract, agar and salts supply essential growth factors such as nitrogen, carbon, sulfurs, minerals and vitamins, particularly the B group. Sodium chloride supplies essential electrolytes like sodium ions for transport and osmotic balance. Bacteriological agar is the solidifying agent.

Formula in g/L

Bacteriological agar	15	Yeast extract	15
Special Peptone Mixture	20	Salts	5

Preparation

Suspend 55 grams of medium in one liter of distilled water. Mix well and dissolve by heating with frequent agitation. Boil for one minute until complete dissolution. Sterilize in autoclave at 121 °C for 15 minutes. Cool to 45-50 °C, mix well and dispense into plates.

Instructions for use

- Inoculate a 15 ml tube of sLB Broth (Cat. 1163) with E. coli sample.

- Incubate at 37 °C for 24 hours in anaerobic conditions.

- Take 10 μl of an aliquot 10^4 cells/ml and inoculate plates of sLB Agar using a Digralsky spreader.

- Incubate at 37 °C overnight.

Quality control

Solubility	Appareance	Color of the dehydrated medium	Color of the prepared medium	Final pH (25°C)
w/o rests	Fine powder	Beige	Amber, slightly opalescent	7,0±0,2

Microbiological test

Incubation conditions: (37 °C / overnight).

Microorganisms

Specification

Escherichia coli DH5 alpha + pUC19

Storage

Temp. Min.:2 °C Temp. Max.:25 °C

sLB Broth

Medium designed to increase bacterial growth and leads to high yields of low copy plasmids and extra high yields of high copy plasmids.

Practical information

Aplications Preparation and recovery of competent cells Categories Escherichia coli

Industry: Culture media for Molecular biology

Principles and uses

sLB Broth has been formulated to significantly increase cellular density when compared to the traditional LB Broth. In the standard LB Broth, E. coli cells reach an abrupt stationary phase upon consumption of nutrients contained in the medium. Cell multiplication is stopped, some cell die and plasmid are lost.

Based on the findings of extensive research, our laboratories have developed a new formulation using a proprietary peptone mixture, yeast extract and salts which allow recombinant E. coli cells to have a higher growth. At the end of the log phase replication continues, thus obtaining higher DNA plasmid yields.

sLB Broth cultures have shown cell stability up to 3 days without cell death, being this one a more convenient medium that eliminates the need of constant attention. E.coli's growth is higher in sLB and buffered sLB Broths than in standard LB after 3 days at 37 °C.

The special peptone mixture, yeast extract and salts supply essential growth factors such as nitrogen, carbon, sulfurs, minerals and vitamins, particularly the B group. Sodium chloride supplies essential electrolytes: Sodium ions for transport and osmotic balance.

Formula in g/L

Yeast Extract, Special peptone Mixture, Salts

Preparation

Suspend 40 grams of the medium in one liter of distilled water. Mix well and dissolve by heating with frequent agitation. Boil for one minute until complete dissolution. Dispense in tubes and sterilize in autoclave at 121 °C for 15 minutes.

Instructions for use

- Carry out the experimental procedure according to appropriate use or purpose.

- Inoculate and incubate at a temperature of 35±2 °C for 24-72 hours.

Quality control

Solubility	Appareance	Color of the dehydrated medium	Color of the prepared medium	Final pH (25°C)
w/o rests	Fine powder	Beige	Amber, slightly opalescent	7,0±0,2



40

Microbiological test

Incubation conditions: (35±2 °C / 24, 48, 72 h).

Microorganisms

Escherichia coli DH5 alpha + pUC19

Storage

Temp. Min.:2 °C Temp. Max.:25 °C Specification Good growth

sLB Broth (Buffered)

Medium designed to increase bacterial growth and leads to high yields of low copy plasmids and extra high yields of high copy plasmids.

Practical information

Aplications

Preparation and recovery of competent cells

Industry: Molecular biology / Microbiological Culture Media

Principles and uses

sLB Broth (Buffered) has been formulated to significantly increase cellular density when compared to the traditional LB Broth. In the standard LB Broth, E. coli cells reach an abrupt stationary phase upon consumption of nutrients contained in the medium. Cell multiplication is stopped, some cell die and plasmid are lost.

Based on the findings of extensive research, our laboratories have developed a new formulation using a proprietary peptone mixture, yeast extract and salts which allow recombinant E. coli cells to have a higher growth. At the end of the log phase replication continues, thus obtaining higher DNA plasmid vields.

sLB Broth cultures have shown cell stability up to 3 days without cell death, being this one a more convenient medium that eliminates the need of constant attention. E.coli's growth is higher in sLB and buffered sLB Broths than in standard LB after 3 days at 37 °C.

The special peptone mixture, yeast extract, agar and salts supply essential growth factors such as nitrogen, carbon, sulfurs, minerals and vitamins, particularly the B group. Sodium chloride supplies essential electrolytes: Sodium ions for transport and osmotic balance.

Formula in q/L

Yeast Extract, Special peptone Mixture, Salts

Preparation

Suspend 54,48 grams of the medium in one liter of distilled water. Mix well and dissolve by heating with frequent agitation. Boil for one minute until complete dissolution. Dispense in tubes and sterilize in autoclave at 121 °C for 15 minutes.

Instructions for use

- Carry out the experimental procedure according to appropriate use or purpose.

- Inoculate and incubate at a temperature of 35±2 °C for 24, 48 and 72 hours.

Quality control

Solubility	Appareance	Color of the dehydrated medium	Color of the prepared medium	Final pH (25°C)
w/o rests	Fine powder	Beige	Amber, slightly opalescent	7,0±0,2

Microbiological test

Incubation conditions: (35±2 °C / 24, 48, 72 h).

Microorganisms

Escherichia coli DH5 alpha + pUC19

Specification Good growth Cat. 1199

54.48

Categories Escherichia coli

Storage

Temp. Min.:2 °C Temp. Max.:25 °C

SOB Medium

For the cultivation of recombinant strains of E.coli

Practical information

Aplications Preparation and recovery of competent cells

Industry: Culture media for Molecular biology

Categories Escherichia coli



Principles and uses

SOB Medium is a nutrient rich medium for the preparation and transformation of competent cells. The transformation requires perforation of the bacteria to allow the introduction of alien DNA inside the cell. In order to survive this process the competent cells need an isotonic rich medium.

Peptone provides nitrogen, vitamins, minerals and amino acids essential for growth. Yeast extract is source of vitamins, particularly the B-group. Sodium chloride and Potassium chloride supplies essential electrolytes for transport and osmotic balance. Magnesium sulfate is a source of magnesium ions.

Formula in g/L

Magnesium chloride anhydrous	0,96	Potassium chloride	0,186
Sodium chloride	0,5	Tryptone	20
Yeast extract	5		

Preparation

Suspend 26,6 grams of the medium in one liter of distilled water .Mix well and dissolve by heating with frequent agitation. Boil for one minute until complete dissolution. Dispense into appropriate containers and sterilize in autoclave at 121°C for 15 minutes.

Instructions for use

Inoculate with the transformed cells and incubate at $35 \pm 2^{\circ}$ C for 18 - 24 hours.

Quality control

Solubility	Appareance	Color of the dehydrated medium	Color of the prepared medium	Final pH (25°C)
w/o rests	Fine powder	Beige	Amber, slightly opalescent	7,0 ± 0,2

Microbiological test

Incubation conditions: (35±2 °C / 18-24 h)

Microorganisms

Escherichia coli ATCC 23724

Specification Good growth

Escherichia coli ATCC 33694 Escherichia coli ATCC 33849 Escherichia coli ATCC 39403 Escherichia coli ATCC 47014

Storage

Temp. Min.:2 °C Temp. Max.:25 °C Good growth Good growth Good growth Good growth

SOC Medium

For the cultivation of recombinant strains of E.coli

Practical information

Aplications

Preparation and recovery of competent cells

Industry: Culture media for Molecular biology

Categories General use





Principles and uses

SOC Medium is a nutrient rich medium for the preparation and transformation of competent cells. The transformation requires perforation of the bacteria to allow the introduction of alien DNA inside the cell. In order to survive this process the competent cells need an isotonic rich medium.

Peptone provides nitrogen, vitamins, minerals and amino acids essential for growth. Yeast extract is source of vitamins, particularly the B-group. Sodium chloride and Potassium chloride supplies essential electrolytes for transport and osmotic balance. Magnesium sulfate is a source of magnesium ions. Glucose is used as a carbon and energy source that E. coli uses to repair the perforation as well as for replication.

Formula in g/L

Glucose 3,6	Magnesium chloride anhydrous	0,96
Potassium chloride 0,186	Sodium chloride	0,5
Tryptone 20	Yeast extract	5

Preparation

Suspend 30,2 grams of the medium in one liter of distilled water. Mix well and dissolve by heating with frequent agitation. Boil for one minute until complete dissolution. Dispense into appropriate containers and sterilize in autoclave at 121 °C for 15 minutes.

Instructions for use

- Inoculate and incubate at 35±2 °C for 18-24 hours.

Quality control

Solubility	Appareance	Color of the dehydrated medium	Color of the prepared medium	Final pH (25⁰C)
w/o rests	Fine powder	Beige	Amber, slightly opalescent	7,0 ± 0,2

Microbiological test

Incubation conditions: (35±2 °C / 18-24 h)

Microorganisms

Specification

Storage

Temp. Min.:2 °C Temp. Max.:25 °C

Super Growth Autoinducible with Trace Elements

To grow IPTG inducible expression in bacterial strains.

Practical information

Aplications Protein expression Categories Escherichia coli

Industry: Molecular biology / Microbiological Culture Media

Principles and uses

Auto induction media was first formulated and developed by W. Studier to grow IPTG-inducible expression strains. The principle of auto induction media is based on carbon sources in the medium that are metabolized differentially to promote high density cell growth and automatically induce protein expression from lac promoters.

Auto induction media contains glucose as well as lactose as the carbon source. A limited concentration of glucose is metabolized preferentially during growth, which prevents uptake of lactose until the glucose is depleted, usually in mid to late log phase. As the glucose is depleted, lactose can be taken up and converted by the enzyme ß-galactosidase to the inducer allolactose. Allolactose causes the release of lac repressor from its specific binding sites in the DNA and thereby induces expression of T7 RNA polymerase from the lacUV5 promoter and unblocks T7lac promoters, allowing expression of target proteins by T7 RNA polymerase.

With Auto induction media, a high density cell growth is followed by a spontaneous induction of protein expression. There is no need to monitor the cell density and there is no conventional induction with IPTG. Parallel growth of many non-induced or auto-induced cultures is feasible because cultures are simply inoculated and grown to saturation. This is a great convenience and simplifies manual or automated induction and analysis of multiple clones compared to conventional IPTG induction, which requires monitoring growth of each culture and adding inducer at the proper stage of growth.

Formula in g/L

Glucose	0,5	Ammonium sulfate	3,3
Disodium phosphate	7,1	Magnesium sulfate	0,15
Monopotassium phosphate	6,8	Tryptone	35
Yeast extract	20	Trace elements	0,015
Alpha lactose	2		

Preparation

Suspend 74,86 grams of the medium in one liter of distilled water. Mix well and dissolve by heating with frequent agitation. Boil for one minute until complete dissolution. Sterilize in autoclave at 115 °C for 20 minutes. Mix well and dispense into appropriate containers.

Instructions for use

- Consult appropriate references for recommended test procedures.

- Incubate at a temperature of 35±2 °C for 18-48 hours.

Quality control

Solubility	Appareance	Color of the dehydrated medium	Color of the prepared medium	Final pH (25ºC)
w/o rests	Fine powder	Beige	Amber	7,0±0,2

Microbiological test

Incubation conditions: (35±2 °C / 18-48 h)

Mucroorc	ianisms
million oon g	garnorno

Escherichia coli ATCC 23724 Escherichia coli ATCC 33694 Escherichia coli ATCC 33849 Escherichia coli ATCC 39403 Escherichia coli ATCC 47014

Storage

Temp. Min.:2 °C Temp. Max.:25 °C Specification Good growth Good growth Good growth Good growth Good growth

Terrific Broth

Medium used with glycerol for the cultivation of recombinant strains of E.coli.

Practical information

 Aplications
 Categories

 Preparation and recovery of competent cells
 Escherichia coli

 Industry: Culture media for Molecular biology
 Escherichia coli



Principles and uses

Terrific Broth is a medium which supports a high cell density and, in the case is formulated for optimum growth of E.coli, maintains growth in the logarithmic phase for a long time. As a result, it yields a greater number of recombinant proteins and plasmic DNA. In some circumstances it substitutes LB Broth (Lennox) (Cat. 1231) used in genetic studies.

Tryptone provide nitrogen, vitamins, minerals and amino acids essential for growth Yeast extract is source of vitamins, particularly the B-group. Potassium phosphates act as a buffer system to prevent cell death.

The source of carbohydrates and carbon is glycerol that is not fermented to acetic acid as glucose and does not lead to confusing results.

Formula in g/L

Dipotassium phosphate	12,54	Monopotassium phosphate	2,31
Tryptone	12	Yeast extract	24

Preparation

Suspend 50.8 grams of the medium in 900 ml of distilled water. Mix well and add 4 ml of glycerol. Adjust to a final volume of 1000 ml and dissolve by heating with frequent agitation. Boil for one minute until complete dissolution. Dispense into appropriate containers and sterilize in autoclave at 121°C for 15 minutes.

Instructions for use

- Carry out the experimental procedure according to appropriate use or purpose.

- Inoculate and incubate at a temperature of 35±2 °C for 18-24 hours.

Quality control

Solubility	Appareance	Color of the dehydrated medium	Color of the prepared medium	Final pH (25°C)
w/o rests	Fine powder	Beige	Amber	7,2±0,2

Microbiological test

Incubation conditions: (35±2 °C / 18-24 h)

Microorganisms	Specification
Escherichia coli ATCC 23724	Good growth
Escherichia coli ATCC 33694	Good growth
Escherichia coli ATCC 33849	Good growth
Escherichia coli ATCC 39403	Good growth
Escherichia coli ATCC 47014	Good growth

Storage

Temp. Min.:2 °C Temp. Max.:25 °C

Terrific Broth (Non animal origin)

Medium used with glycerol for the cultivation of recombinant strains of E.coli.

Practical information

Aplications Protein expression Categories Escherichia coli

Industry: Culture media for Molecular biology



Principles and uses

Terrific Broth (Non animal origin) is as Terrific Broth (Cat. 1246) but changing the animal origin raw materials to vegetal origin ones. It is a medium which supports a high cell density and, in the case is formulated for optimum growth of E.coli, maintains growth in the logarithmic phase for a long time. As a result it yields a greater number of recombinant proteins and plasmic DNA. In some circumstances it substitutes LB Broth (Lennox) (Cat. 1231) used in genetic studies.

Vegetable origin peptone provide nitrogen, vitamins, minerals and aminoacids essential for growth. Yeast extract is source of vitamins, particularly the B-group. Potassium phosphates act as a buffer system to prevent cell death.

Formula in g/L

Dipotassium phosphate	12,5	Monopotassium phosphate	2,3
Yeast extract	24	Vegetable peptone	12

Preparation

Suspend 50,8 grams of the medium in 1 liter of distilled water. Mix well and dissolve by heating with frequent agitation. Boil for one minute until complete dissolution. Dispense into appropriate containers and sterilize in autoclave at 121 °C for 15 minutes.

Instructions for use

Inoculate and incubate at 35±2 °C for 18-24 hours.

Quality control

Solubility	Appareance	Color of the dehydrated medium	Color of the prepared medium	Final pH (25ºC)
w/o rests	Fine powder	Beige	Amber	7,2±0,2

Microbiological test

Incubation conditions: (35±2 °C / 18-24 h).

Microorganisms

Specification

Escherichia coli ATCC 23724 Escherichia coli ATCC 33694 Escherichia coli ATCC 33849 Escherichia coli ATCC 39403 Escherichia coli ATCC 47014

Storage

Temp. Min.:2 °C Temp. Max.:25 °C Good growth Good growth Good growth Good growth Good growth

Terrific Broth Base Autoinducible w/o Metals

For the expression of inducible IPTG bacterial strains.

Practical information

Aplications Protein expression Categories Escherichia coli

Industry: Culture media for Molecular biology

Principles and uses

Terrific Broth Base Autoinducible w/o Metalsis a medium which supports a high cell density and, in the case is formulated for optimum growth of E.coli, maintains growth in the logarithmic phase for a long time. As a result, it yields a greater number of recombinant proteins and plasmic DNA. In some circumstances, it substitutes LB Broth (Lennox) (Cat. 1231) used in genetic studies.

Auto Induction Media was firstly formulated and developed by W. studier to grow IPTG-inducible expression strains. The principle of Auto Induction Media is based on carbon sources in the medium that are metabolized differentially to promote high density cell growth and automatically induce protein expression from lac promoters. Auto Induction Media contains glucose as well as lactose as the carbon source. A limited concentration of glucose is metabolized preferentially during growth, which prevents uptake of lactose until the glucose is depleted, usually in mid to late log phase. As the glucose is depleted, lactose can be taken up and converted by the enzyme ß-galactosidase to the inducer allolactose. Allolactose causes the release of lac repressor from its specific binding sites in the DNA and thereby induces expression of T7 RNA polymerase from the lacUV5 promoter and unblocks T7lac promoters, allowing expression. There is no need to monitor the cell density and there is no conventional induction with IPTG. Parallel growth of many non-induced or auto-induced cultures is feasible because cultures are simply inoculated and grown to saturation. This is a great convenience and simplifies manual or automated induction and analysis of multiple clones compared to conventional IPTG induction, which requires monitoring growth of each culture and adding inducer at the proper stage of growth. For higher saturation density use the Super Growth Autoinducible Medium w/o Trace Element (Cat. 2095).

Tryptone provide nitrogen, vitamins, minerals and amino acids essential for growth. Yeast extract is source of vitamins, particularly the B-group. Potassium phosphates act as a buffer system to prevent cell death. If desired, add a concentration of 0.015 g/l of Trace Elements (Cat. 2109) to supply all of the specific requirements for the bacteria.

Formula in g/L

Glucose	0,5	Alpha-lactose	2
Ammonium sulfate	3,3	Disodium phosphate	7,1
Magnesium sulfate 0	,15	Monopotassium phosphate	6,5
Tryptone	12	Yeast extract	24

Preparation

Suspend 55,55 grams of the medium in one liter of distilled water. Mix well and dissolve by heating with frequent agitation. Boil for 1 minute or until complete dissolution. Sterilize in autoclave at 115 °C for 20 minutes. Mix well and dispense as wished.

Instructions for use

Inoculate and incubate at a temperature of 35±2 °C for 18-48 hours.



Quality	control	

Solubility	Appareance	Color of the dehydrated medium	Color of the prepared medium	Final pH (25°C)
w/o rests	Fine powder	Beige	Amber	7,0±0,2

Microbiological test

Incubation conditions: (35±2 ° / 18-48 h).

Microorganisms	Specification
Escherichia coli ATCC 23724	Good growth
Escherichia coli ATCC 33694	Good growth
Escherichia coli ATCC 33849	Good growth
Escherichia coli ATCC 39403	Good growth
Escherichia coli ATCC 47014	Good growth

Storage

Temp. Min.:2 °C Temp. Max.:25 °C

Vegetable LB Agar (Lennox)

Recommended medium for maintaining and cultivating recombinant strains of E. coli.

Practical information

Aplications

Preparation and recovery of competent cells

Categories Escherichia coli

Industry: Molecular biology / Microbiological Culture Media

Principles and uses

Vegetable LB Agar (Lennox) is a nutritionally rich medium based on the LB Agar (Lennox) (Cat. 1083) designed as an alternative to classical animal-based media for the growth and maintenance of pure cultures of recombinant strains of E. coli used in molecular microbiology procedures.

These strains are generally derived from E. coli K12, which are unable to produce vitamin B, so this media is formulated to enhance the growth of nutritionally demanding microorganisms. This strain of E. coli has been further modified through specific mutation to create an auxotrophic strain that is not capable of growth on nutritionally deficient media.

In this case, the ingredients are animal-free in order to minimize the risk of bovine spongiform encephalopathy in culture media containing bovine materials. Vegetable origin peptone provides nitrogen, vitamins, minerals and amino acids essential for growth. Yeast extract is source of vitamins, particularly the B-group essential for bacterial growth. Sodium chloride supplies essential electrolytes for transport and osmotic balance. Bacteriological agar is the solidifying agent.

Vegetable LB Agar (Lennox) has a different sodium chloride level than other media such as Luria Agar (Miller LB Agar) (Cat. 1552) or Luria Agar (Miller's Modification) (Cat. 1308). This allows to select the optimum salt concentration of the medium for a specific strain.

Formula in g/L

Bacteriological agar 1	15 S	Sodium chloride	5
Yeast extract	5 \	Vegetable peptone	10

Preparation

Suspend 35 grams of medium in one liter of distilled water. Mix well and dissolve by heating with frequent agitation. Boil for one minute until complete dissolution. Sterilize in autoclave at 121 °C for 15 minutes. Cool to 45-50 °C, mix well and dispense into plates.

Instructions for use

- Carry out the experimental procedure according to appropriate use or purpose.

- Inoculate and incubate at a temperature of 35±2 °C for 18-24 hours.

Quality control

Solubility	Appareance	Color of the dehydrated medium	Color of the prepared medium	Final pH (25°C)
w/o rests	Fine powder	Beige	Amber, slightly opalescent	7,0±0,2

Microbiological test

Incubation conditions: (35±2 °C / 18-24 h).

Microorganisms

Escherichia coli ATCC 23724

Specification Good growth

Escherichia coli ATCC 33694 Escherichia coli ATCC 33849 Escherichia coli ATCC 39403 Escherichia coli ATCC 47014

Storage

Temp. Min.:2 °C Temp. Max.:25 °C Good growth Good growth Good growth Good growth

Vegetable LB Broth (Lennox)

Recommended medium for maintaining and cultivating recombinant strains of E. coli.

Practical information

Aplications Preparation and recovery of competent cells Categories Escherichia coli

Industry: Culture media for Molecular biology

Principles and uses

Vegetable LB Broth (Lennox) is a nutritionally rich medium based on the LB Broth (Lennox) (Cat. 1163) designed as an alternative to classical animal-based media for the growth and maintenance of pure cultures of recombinant strains of E. coli used in molecular microbiology procedures.

These strains are generally derived from E. coli K12, which are unable to produce vitamin B, so this media is formulated to enhance the growth of nutritionally demanding microorganisms. This strain of E. coli has been further modified through specific mutation to create an auxotrophic strain that is not capable of growth on nutritionally deficient media.

In this case, the ingredients are animal-free in order to minimize the risk of bovine spongiform encephalopathy in culture media containing bovine materials. Vegetable origin peptone provides nitrogen, vitamins, minerals and amino acids essential for growth. Yeast extract is source of vitamins, particularly the B-group essential for bacterial growth. Sodium chloride supplies essential electrolytes for transport and osmotic balance.

Vegetable LB Broth (Lennox) has a different sodium chloride level than other media such as Luria Broth (Miller LB Broth) (Cat. 1551) or Luria Broth (Miller's Modification) (Cat. 1266). This allows to select the optimum salt concentration of the medium for a specific strain.

Formula in g/L

Sodium chloride	5	Yeast extract	5
Vegetable peptone	10	_	_

Preparation

Suspend 20 grams of the medium in one liter of distilled water. Mix well and dissolve by heating with frequent agitation. Boil for one minute until complete dissolution and dispense into tubes. Sterilize in autoclave at 121 °C for 15 minutes.

Instructions for use

Carry out the experimental procedure according to appropriate use or purpose.
 Inoculate and incubate at a temperature of 35±2 °C for 18-24 hours.

Quality control

Solubility	Appareance	Color of the dehydrated medium	Color of the prepared medium	Final pH (25°C)
w/o rests	Fine powder	Beige	Clear beige	7,0±0,2

Microbiological test

Incubation conditions: (35±2 °C / 18-24 h).

Microorganisms	Specification
Escherichia coli ATCC 23724	Good growth
Escherichia coli ATCC 33694	Good growth
Escherichia coli ATCC 33849	Good growth
Escherichia coli ATCC 39403	Good growth
Escherichia coli ATCC 47014	Good growth

Storage

Temp. Min.:2 °C Temp. Max.:25 °C

Yeast Nitrogen Base w/o Added Aminoacids and w/o Ammonium Sulfate

For yeast classification based on carbon and nitrogen requirements.

Practical information

 Aplications
 Categories

 Differentiation
 Yeasts

Industry: Culture media for Molecular biology

Principles and uses

Yeast Nitrogen Base w/o Added Aminoacids and w/o Ammonium Sulfate is used for classifying yeasts based on carbon and nitrogen requirements and is prepared according to the formulas of Wickerharm and Burkholder. The medium contains all the essential vitamins and inorganic salts needed to cultivate yeasts, except for the aminoacids and carbohydrate sources.

This medium is used in many applications for the study of yeast in molecular biology as is useful for the determination of aminoacids and carbohydrate utilization.

Formula in g/L

Biotin	0,000002	Boric acid	0,0005
Calcium chloride	0,1	Calcium patothenate	0,0004
Ferric chloride	0,0002	Folic Acid	0,00002
Inositol	0,002	Magnesium sulfate	0,5
Manganase sulfate	0,0004	Monopotassium phosphate	1
Niacin	0,0004	P-Aminobenzoic acid	0,0002
Potassium iodide	0,0001	Riboflavin	0,0002
Sodium chloride	0,1	Sodium molybdate	0,0002
Thiamine hydrochloride	0,0004	Zinc sulfate	0,0004
Cupric Sulphate	0,00004		

Typical formula g/L * Adjusted and/or supplemented as required to meet performance criteria.

Preparation

Prepare a 10X solution by dissolving 1,7 grams of the medium in 100 ml of distilled water with 5 grams of dextrose, or the equivalent amount of another carbohydrate, and 5-10 mg of the desired amino acid. Mix well. Heat with frequent agitation until complete dissolution. DO NOT BOIL. DO NOT AUTOCLAVE. Sterilize the solution by filtration. Prepare the final medium by aseptically pipetting 0,5 ml of the 10X solution to 4,5 ml of distilled water. Swirl to mix solution before inoculation.

Instructions for use

Inoculate and incubate at a temperature of 25-30 °C for 2-5 days.

Quality control

Solubility	Appareance	Color of the dehydrated medium	Color of the prepared medium	Final pH (25°C)
w/o rests	Fine powder	Off-white	Colorless	4,5±0,2

Microbiological test

Incubation conditions: (25-30 °C / 2-5 days).

Microorganisms

Candida albicans ATCC 10231 Sacharomyces cerevisiae ATCC 9080 Saccharomyces cerevisiae ATCC 9763 Kloeckera apiculata ATCC 9774

Storage

Temp. Min.:2 °C Temp. Max.:25 °C Specification Good growth Good growth Good growth

Yeast Nitrogen Base w/o Amino Acids

For yeast classification based on amino acids and carbohydrates requirements

Practical information

Aplications Differentiation Categories Yeasts

Industry: Food / Culture media for Molecular biology

Principles and uses

Yeast Nitrogen Base w/o Amino Acids is used for classifying yeasts based on carbon and nitrogen requirements and is prepared according to the formulas of Wickerharm and Burkholder. The medium contains all the essential vitamins and inorganic salts needed to cultivate yeasts, except for the amino acids and carbohydrate sources. Ammonium sulphate is included as a readily available nitrogen source for nitrogen assimilation.

This medium is used in many applications for the study of yeast in molecular biology as is useful for the determination of aminoacids and carbohydrate utilization.

Formula in g/L

Ammonium sulfate	5	Boric acid	0,0005
Calcium chloride	0,1	Calcium patothenate	0,0004
Ferric chloride	0,0002	Inositol	0,002
Magnesium sulfate	0,5	Manganase sulfate	0,0004
Monopotassium phosphate	1	Niacin	0,0004
P-Aminobenzoic acid	0,0002	Potassium iodide	0,0001
Pyridoxine hydrochloride	0,0004	Riboflavin	0,0002
Sodium chloride	0,1	Sodium molybdate	0,0002
Thiamine hydrochloride	0,0004	Zinc sulfate	0,0004
Copper sulfate (mg)	0,04	Folic acid (mg)	0,002
Biotin (mg)	0,002		

Preparation

Prepare a 10X solution by dissolving 6,7 grams of the medium in 100 ml of distilled water with 5 grams of dextrose, or the equivalent amount of another carbohydrate, and 5-10 mg of the desired amino acid. Mix well. Heat with frequent agitation until complete dissolution. DO NOT BOIL. DO NOT AUTOCLAVE. Sterilize the solution by filtration. Prepare the final medium by aseptically pipetting 0,5 ml of the 10X solution to 4,5 ml of distilled water. Swirl to mix solution before inoculation.

Instructions for use

- Inoculate de prepared tubed medium very lightly with the test organism.

- Incubate at 25-30 °C for 2-5 days.
- After incubation, shake the tubes to suspend growth.
- Read for growth.

- Carry out the carbon and nitrogen assimilation tests described in reference manuals.



Solubility	Appareance	Color of the dehydrated medium	Color of the prepared medium	Final pH (25°C)
w/o rests	Fine powder	Off-white	Ambar	5,4±0,2
Microbiol	ogical test			
Incubation co	nditions: (25-30 °C / 2-5	days).		
Microorganisn	ns		Specification	
Candida albicans ATCC 10231			Good growth	
Hanseniaspora uvarum ATCC 32856			Good growth	
Sacharomyce	s cerevisiae ATCC 9080		Good growth	

Storage

Temp. Min.:2 °C Temp. Max.:25 °C

YP Agar Base Medium

For maintaining and developing yeast in molecular biology procedures.

Practical information

Aplications Categories Growth Yeasts

Industry: Molecular biology / Microbiological Culture Media

Principles and uses

YP Agar Base Medium is used for the maintenance and the development of yeast in molecular biology procedures.

YP Agar Base Medium is also used to cultivate Saccharomyces cerevisiae and other yeasts. Yeasts grow well on a medium containing only a minimal amount of glucose and salts. This medium contains glucose (with the addition of dextrose after autoclaving), salts and proteins, which favours the growth of Saccharomyces cerevisiae and reduces growing times. Yeast extract is the source of vitamins, particularly the B-group, essential for bacterial growth. Peptone provides nitrogen, vitamins, minerals and amino acids. Bacteriological agar is the solidifying agent.

Saccharomyces cerevisiae has a genome of 14 Mb containing 6.000 genes arranged in 16 chromosomes, which have been completely sequenced, and thus, is a species type in microbiology and genetics studies.

Formula in g/L

Bacteriological agar	20	Peptone	20
Yeast extract	10		

Preparation

Suspend 50 grams of the dehydrated medium in 900 ml of distilled water. Mix well and dissolve by heating with frequent agitation. Boil for one minute until complete dissolution. Sterilize in autoclave at 121 °C for 15 minutes. Cool to room temperature and aseptically add 100 ml of sterile dextrose at 20%. Mix well and dispense into appropriate containers.

Instructions for use

Poured plate method:

- Deposit 1 ml of the initial suspension and/or diluted sample in an empty Petri dish.
- Add 12-15 ml of agar cooled to 44-47 °C in each Petri dish and mix gently moving the plate.
- Allow the plates to solidify and incubate in an inverted position at a temperature of 25±2 °C for 42-48 hours.

Streak plate method:

- In a Petri dish, add 12-15 ml of molten agar and let it solidify.

- Inoculate 10 μl of the initial suspension and/or diluted sample.

- Extend the inoculum with a sterile loop on the agar surface.

- Incubate the plates in an inverted position at a temperature of 25±2 °C for 42-48 hours.

Quality control

Solubility	Appareance	Color of the dehydrated medium	Color of the prepared medium	Final pH (25°C)
w/o rests	Fine powder	Beige	Clear amber, slightly opalescent	7,0±0,2

Microbiological test

Incubation conditions: (25±2 °C / 42-48 h).

Microorganisms Candida albicans ATCC 10231 Saccharomyces cerevisiae ATCC 9763

Storage

Temp. Min.:2 °C Temp. Max.:25 °C Specification Good growth Good growth

YP Base Medium

For maintaining and developing yeast in molecular biology procedures.

Practical information

 Aplications
 Categories

 Growth
 Yeasts

Industry: Molecular biology / Microbiological Culture Media

Principles and uses

YP Base Medium is used for the maintenance and the development of yeast in molecular biology procedures.

YP Base Medium is also used to cultivate Saccharomyces cerevisiae and other yeasts. Yeasts grow well on a medium containing only a minimal amount of glucose and salts. This medium contains glucose (with the addition of dextrose after autoclaving), salts and proteins, which favors the growth of Saccharomyces cerevisiae and reduces growing times. Yeast extract is the source of vitamins, particularly the B-group essential for bacterial growth. Peptone provides nitrogen, vitamins, minerals and amino acids.

Saccharomyces cerevisiae has a genome of 14 Mb containing 6.000 genes arranged in 16 chromosomes, which have been completely sequenced, and thus, is a species type in microbiology and genetics studies.

Formula in g/L

Peptone

20 Yeast extract

10

Cat. 1511

Preparation

Suspend 30 grams of the medium in 900 ml of distilled water. Mix well and dissolve by heating with frequent agitation. Boil for one minute until complete dissolution. Sterilize in autoclave at 121 °C for 15 minutes. Cool to room temperature and aseptically add 100 ml of sterile dextrose at 20 %. Mix well and dispense intro appropriate containers.

Instructions for use

Inoculate and incubate at a temperature of 25±2 °C for 42-48 hours.

Quality control

Solubility	Appareance	Color of the dehydrated medium	Color of the prepared medium	Final pH (25°C)
w/o rests	Fine powder	Beige	Amber, slightly opalescent	6,5±0,2

Microbiological test

Incubation conditions: (25±2 °C / 42-48 h).

Microorganisms	Specification	
Candida albicans ATCC 10231	Good growth	
Saccharomyces cerevisiae ATCC 9763	Good growth	

Storage

Temp. Min.:2 °C Temp. Max.:25 °C



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YPD Agar

For maintaining and developing yeast in molecular biology procedures

Practical information

Aplications Enrichment Categories Yeasts

Industry: Molecular biology / Microbiological Culture Media



Principles and uses

YPD Agar is used for maintaining and developing yeast in molecular microbiology procedures. The formula is the same as in YP Agar Base Medium (Cat. 1513) but with the dextrose added.

YPD Agar is also used to cultivate Saccharomyces cerevisiae and other yeasts. Yeasts grow well on a medium containing only a minimal amount of glucose and salts. This medium contains dextrose (with the addition of dextrose after autoclaving), salts and proteins, which favors the growth of Saccharomyces cerevisiae and reduces growing times. Yeast extract is the source of vitamins, particularly the B-group. Peptone provides nitrogen, vitamins, minerals and amino acids. Bacteriological agar is the solidifying agent.

Saccharomyces cerevisiae has a genome of 14 Mb containing 6000 genes arranged in 16 chromosomes, which have been completely sequenced, and thus, is a species type in microbiology and genetics studies.

Formula in g/L

Bacteriological agar 15	Dextrose	20
Peptone 20	Yeast extract	10

Preparation

Suspend 65 grams of medium in one liter of distilled water. Mix well and dissolve by heating with frequent agitation. Boil for one minute until complete dissolution. Sterilize in autoclave at 121°C for 15 minutes. Cool to 45-50°C, mix well and dispense into plates.

Instructions for use

- This medium can be inoculated directly or after enrichment broth YPD Broth (Cat. 1547).

- Spread the plates of YPD Agar and incubate at 25±2°C for 42-48 hours.

Quality control

Solubility	Appareance	Color of the dehydrated medium	Color of the prepared medium	Final pH (25°C)
w/o rests	Fine powder	Beige	Amber, slightly opalescent	6,5 ± 0,2

Microbiological test

Microorganisms

Candida albicans ATCC 10231 Sacharomyces cerevisiae ATCC 9080

Storage

Temp. Min.:2 °C Temp. Max.:25 °C Specification Good growth Good growth
YPD Broth

For maintaining and developing yeasts in molecular biology procedures.

Practical information

Aplications Enrichment Protein expression

Industry: Culture media for Molecular biology

4

Principles and uses

YPD Broth is used for the maintenance and the development of yeast in molecular biology procedures. The formula is the same as in YP Base Medium (Cat. 1511) but with the dextrose added.

Categories Yeasts

Yeasts

YPD Broth is also used to cultivate Saccharomyces cerevisiae and other yeasts. Yeasts grow well on a medium containing only a minimal amount of glucose and salts. This medium contains dextrose, salts and proteins, which favors the growth of Saccharomyces cerevisiae and reduces growing times. Yeast extract is the source of vitamins, particularly the B-group essential for bacterial growth. Peptone provides nitrogen, vitamins, minerals and amino acids.

Saccharomyces cerevisiae has a genome of 14 Mb containing 6000 genes arranged in 16 chromosomes, which have been completely sequenced, and thus, is a species type in microbiology and genetics studies.

Formula in g/L

Dextrose	20 Peptone	20
Yeast extract	10	

Preparation

Suspend 50 grams of the medium in one liter of distilled water .Mix well and dissolve by heating with frequent agitation. Boil for one minute until complete dissolution. Dispense into appropriate containers and sterilize in autoclave at 118°C for 15 minutes. Avoid overheating.

Instructions for use

- Carry out the experimental procedure according to appropriate use or purpose.

- Inoculate and incubate at a temperature of 25±2 °C for 42-48 hours.

Quality control

Solubility	Appareance	Color of the dehydrated medium	Color of the prepared medium	Final pH (25°C)
w/o rests	Fine powder	Beige	Amber, slightly opalescent	6,5 ± 0,2

Microbiological test

Cat. 1547



Incubation conditions: (25±2 °C / 42-48 h)

Microorganisms

Candida albicans ATCC 10231 Sacharomyces cerevisiae ATCC 9080 Saccharomyces cerevisiae ATCC 9763

Storage

Temp. Min.:2 °C Temp. Max.:25 °C Specification Good growth Good growth

Good growth

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